INTERIOR HORT ... for interiorscape professionals

Center for Urban Horticulture
University of Washington

Cooperative Extension
Washington State University

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INTERIORSCAPE PLANT FORUM: Tough Plants for Tough Locations

Date : Wednesday, February 17
Time : 7 to 9 p.m.
Location : Center for Urban Horticulture
Discussion Leaders : George Pinyuh, Washington State University Cooperative Extension; Toni Pietromonaco, Interior Plant Design; Tim Savage, Growing Green Gardens Inc.; and Sharon Thorsen, Interiors in Green.

Here is a chance to meet with fellow interiorscapers and share information on selecting plants for difficult locations. In the first hour, small groups will discuss special problems—low light, cold and drafty lobbies, high traffic areas, pools and saunas, dirty air, paint fumes, poor air circulation, and extreme temperature fluctuations—encountered by interiorscapers. During the second hour, the groups willl present their results to the entire audience for further discussion. Afterwards, notes from the forum will be typed and sent to all participants. Coffee, tea, and cookies will be served.

Registration Form: Interior Plant Forum

Registration Fee
Before February 10 . . . . . $5.00
After February 10 . . . . . $7.50

Firms using purchase orders must make prior registration arrangements.

Make checks payable to the University of Washington; no bank cards.

Receipts will be available at the door; they will not be returned by mail.

NAME_____

ADDRESS_____

CITY_____

STATE_____ZIP_____

PHONE (DAY)_____

PHONE (EVE)_____

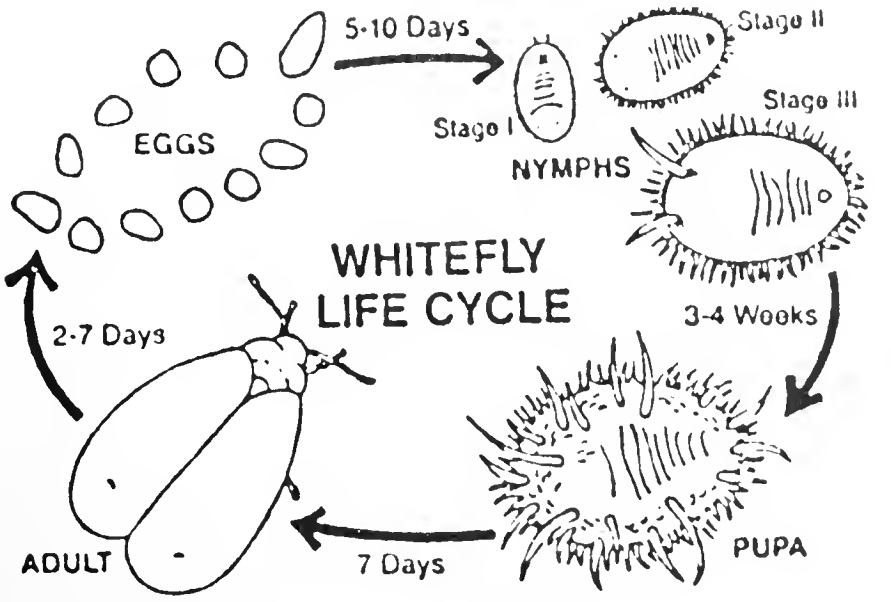
Mail payment and registration to: Urban Horticulture Program, University of Washington, GF–15, Seattle, WA 98195

For more information, please call 545–8033.

The Whitefly Life Cycle

Greenhouse whiteflies are small white insects that resemble miniature moths. The adults are commonly found on the underside of leaves and fly for a short distance when disturbed. They can infest a wide range of crops anytime of the year.

Each adult female can lay up to 200 eggs in her life time. Normally the females lay up to 20 eggs per batch in a circle on the underside of a leaf. Tiny crawlers emerge from the eggs in 5 to 10 days and seek a feeding site. They insert their piercing, sucking mouth parts into the leaf tissue and remain stationary for 3 to 4 weeks. The flat scab-like green to yellowish-colored transparent nymphs undergo 3 nymphal molts and then pupate. They remain in the pupal stage approximately 1 week after which they emerge as adults. Females then begin laying eggs 2 to 7 days later. Depending upon temperature, the whitefly's life cycle (egg to adult) can be completed in 5 to 7½ weeks. All feeding stages (nymphs and adults) of the whitefly have piercing, sucking mouth parts.



* This article was reprinted from the *Illinois State Florists' Association Bulletin*, May–June 1987, and originally appeared in the *Erie County (NY) Horticultural Notes*, December 1986.

Do Leaf Shines Reduce Low Light Tolerance?

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Foliage plant growers commonly spray their plants with leaf shine materials prior to shipping. The leaf shine cleans residues off the foliage and makes the plants more attractive. But research reported by the University of Florida in 1983 shows that *Ficus benjamina* is less tolerant of low light conditions when treated with leaf shine prior to shipment. Two experiments tested the effects of four commercial leaf shine products—Volck oil spray, Green-Glo, Foliage Plant Leaf Polish, and Luster Leaf—on the tolerance of *Ficus benjamina* to low light conditions.

The light compensation point (LCP) was used to measure low light tolerance. Plants with a lower LCP have a greater tolerance of low light conditions. For example, it has been shown that the LCP of *Ficus benjamina* can be lowered by increasing shade levels and decreasing fertilization during production; such acclimated plants can be moved into interiorscapes with minimal leaf drop.

Table 1. Influence of leaf shine compounds on light compensation point of *Ficus benjamina*.

Treatment	Light compensation point (μmol) Expt. 1 (1979)	Expt. 2 (1980)
Control (No spray)	28.4 a[z]	40.8 a
Oil Spray (Volck. 1%)	38.0 b	56.2 c
Green-Glo (no dilution)	36.2 b	52.5 bc
Foliage Plant Leaf Polish (diluted 1:30)	34.2 b	47.3 ab
Luster Leaf (diluted 1:9)	34.6 b	51.0 bc

[z] Mean separation within treatments by Duncan's multiple range test, 5% level.

In both experiments, the plants were grown for five months. Then the upper surfaces of the leaves were sprayed with leaf shine. After being sprayed, the plants were placed in a dark room for five days to simulate transportation to market. The plants were then removed and their LCPs measured.

As shown in Table 1, plants treated with leaf shine materials had higher LCPs than the control plants in both experiments. The reflective quality of the leaf shines may cause an increase in LCP. If so, higher light levels would be required for the shine-treated plants to photosynthesize at the same rate as the control plants.

According to the researchers: "Treated plants lost about 3 times more leaves than did the controls within 2 weeks and, thus, the results indicate a need to increase light in interiorscapes to maintain quality of shine-treated plants compared with non-treated ones."

Reference: Joiner, J.N., C.A. Conover, and R. T. Poole. 1983. Influence of leaf shine compounds on light compensation point of *Ficus benjamina*. HortScience 18:373–374.

Crown and Root Rot of African Violet

According to R. K. Jones and D. L. Strider of North Carolina State University:

> Crown and root rot caused by the fungus *Phytophthora parasitica* is the most serious disease of African violet. This disease occurs at all stages of production from propagation to marketing. The fungus moves from the roots and crowns into the petioles and leaves. Phytophthora-infected tissue is dark brown with a water soaked appearance. Diseased plants wilt and die quickly.
>
> Disease development is favored by warm temperatures and excess water. The disease can be a very serious problem in subirrigation using a sand base. The fungus has not been reported to spread in capillary mat watering systems.

Reference: Jones, R. K., and D. L. Strider. 1986. African violet diseases. Illinois State Florists' Association Bulletin No. 424.

Relative Susceptibility of African Violet to Phytophthora Root and Crown Rot.

Susceptibility	Cultivar: Rhapsody Series	Ballet Series	Melodie Series	Optimara Series
Highly Resistant	Barbara Astrid Ruby	Erica Inge Helga Karla	Kathy	New York California
Resistant	Elfriede Pluto Linda Michele Mars Cornelia Veronika Apollo Audrey	Dolly Anna Rachel	Allison Frances Pearl Diane Angie Stacy	Maryland
Susceptible	Denise Gigi Venus Ruth Gloria Bettina	Ulli Heidi Cristina Eva Meta	Farrah Beth Ellen Julianne	Colorado Virginia
Highly Susceptible	Maria Sophie Mercury Jupiter Neptune Candy Vanessa	Annette Marta Lisa Apollo Carmen	Sheri Mitzi Irene Adeline Suzanne	